Mode of action of a multi-microbial inoculant on pathogen suppression and impact on soil microbial communities





Introduction

Recently, a new product was introduced with potential to control plant diseases. The multi-microbial inoculant (MI) product consists of numerous species of benign and probiotic microorganisms that are cocultured in an aqueous medium. The MI product mediates diverse activities including: (1) suppressing plant pathogen activity and (2) enhancing microbial activity (i.e., decomposition of crop residues). Modes of action driving these activities are unknown. We hypothesize that the modes of action contributing to microbial efficacy include: (1) the enhancement of soil microbial community diversity and (2) interference with pathogen quorum sensing (QS) pathways.

We present an approach for testing these hypotheses and share preliminary results. The first hypothesis is approached by analyzing microbial DNA samples from soils treated with MI. To test the second hypothesis we examine possible breakdown of QS autoinducers (AI) upon exposure to MI using Gas Chromatography-Mass Spectrometry (GC-MS).

Methods

Objective 1

Analyze the diversity of soil microbial communities after MI application to soil.

<u>Approaches</u>

MI was applied to soils in the field and intact soil microcosms incubated in a growth chamber.

1. Field Setting

- Land Management (Fig. 1):
- Cultivated Soil or
- Reintroduced Grassland

Treatments:

- Three spray applications of MI (100 L ha⁻¹) or
- Three spray applications of H₂O (100 L ha⁻¹) (control)
- **Replication:**

- Each land management x treatment is replicated in triplicate (12 plots total)

Plot dimensions: 3m x 5m

Study initiated in March 2012 and plots were sampled eight times between initiation and **October 2012.**

2. Growth Chamber Study of Intact Soil <u>Microcosms</u>

Soil microcosms:

- 15 intact soil cores (ISC) were obtained from cultivated soil control plots and planted to tomato (Solanum lycopersicum)

Treatments (MI inoculation rates and dates)

- 40L ha⁻¹ (April) and 3x20L ha⁻¹ (May-October)
- 40L ha⁻¹ (April) and 60L ha⁻¹ (May)
- 40L ha⁻¹ (April) and 60L ha⁻¹ (July)
- 60L ha⁻¹ (October)

- Control received corresponding amounts of H₂O whenever MI was applied

Replication:

- Each treatment and control is replicated in triplicate (15 ISC total)

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Methods (continued)

Objective 2

Monitor changes in QS autoinducer (AI) concentrations during exposure to MI or H_2O .

<u>Approach (Fig. 2.)</u>

- Two Als examined
- N-(3-Oxo<u>hexa</u>noyl)-L-homoserine lactone (AHLEc) 2. N-(3-Oxodecanoyl)-L-homoserine lactone (AHL)
- Standard curves for AI were developed using GC-MS and used for sample analyses
- Als were exposed to MI or H_2O for 2, 4, and 8 hours
- After exposure, Als were extracted by liquid-liquid extraction with chloroform
- Chloroform extracts were analyzed using a GC-MS (Fig. 2) to determine AI concentrations

Figure 1. Experimental Approach (Objective 1)









Intact Soil Cores in **Growth Chambe**





Soil Sampling

And Soil DNA Extraction

Visual (software enhanced) analysis of DGGE images









Fig. 3. Chromatograms of AHL extracts; 2h exposure to water (red)

Results support the hypothesis that MI affect QS Autoinducers. However, AI degradation appears to be not the only indicator of MI impact; MI may rapidly convert AH to AHLA for subsequent degradation.



N-(3-Oxodecanoyl)-L-homoserine (Peak B) (Fig. 4).

•AHLEc analysis is in progress

Preliminary Conclusions

Acknowledgments

We thank to Dr. Korsi Dumenyo who provided necessary supplies and to Margarita Correa, Sławomir Gacka, Stanisław Kolbusz, Narin Tipsrisukond, Matthew Wood for their guidance and comments. Funding was provided by SCD Probiotics, LLC and Probiotics Polska.