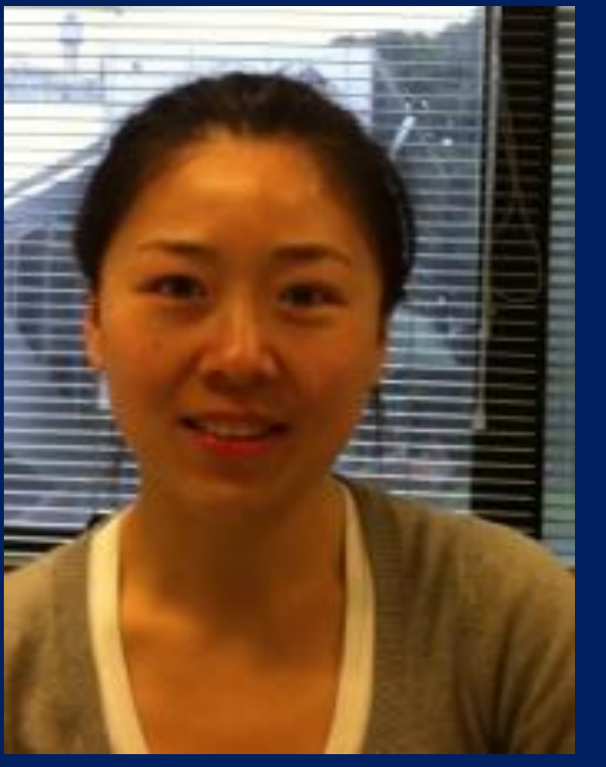


How Do Soil Properties Affect the Efficacy of Nitrification Inhibitors On Nitrous Oxide Emission and Distribution of Ammonia Oxidizers?



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Introduction

Nitrous oxide (N₂O) is a major GHG produced by agricultural practices. Denitrification and nitrification are the two main N₂O producing processes in soils. The application of nitrification inhibitors is one strategy to reduce N₂O losses as they inhibit the bacterial ammonia monooxygenase (AMO) enzyme that is involved in the oxidation of NH₃ to NH₂OH, the first step of nitrification (Fig. 1). However, previous studies have indicated that the efficacy of nitrification inhibitors is variable both spatially and temporally, and this depends greatly on the environmental conditions and soil properties. It has been observed that application of fertilizers treated with nitrification inhibitors had impact on soil ammonia-oxidizing bacterial (AOB) populations but not on ammonia oxidizing-archaeal (AOA) populations (Di et al., 2010; O'Callaghan et al., 2010). However, recent results have indicated that nitrification inhibitors can stop AOA growth (Zhang et al., 2012). Differences in soil properties seems to be a key parameter responsible for the variation in AOA & AOB distributions.

Objective

To investigate how soil physical, chemical and microbial (AOB and AOA abundance) properties influence the efficacy of the nitrification inhibitors 3,4-dimethylpyrazole phosphate (DMPP) and acetylene (C₂H₂) in reducing nitrification and N₂O production.

Materials and Methods

- ❖ Laboratory Incubation Experiment
- ❖ Soils: See Table 1
- ❖ Treatment: NH₄Cl (100 µg NH₄-N/g soil (for all treatments)) (Control)
 NH₄Cl + DMPP (0.1% active ingredient) (DMPP)
 NH₄Cl + C₂H₂ (1% v/v) (C₂H₂)

4 replications at 25 °C and 60% water-filled pore space (WFPS)

❖ Measurements:

- N₂O – flux measurements taken over 72 hrs on days 1, 4, 8 and 12, analysed using gas chromatograph.
- Soil mineral N – extracted weekly with 2M KCl, analysed by segmented-flow analyser (Skalar SAN+++)
- Gene Abundance - DNA extraction (DNA Isolation Kit (MOBIO Laboratories, Inc., US)) → cloning (TOPO cloning kit) → sequencing → quantifying (qPCR)

Table 1. Selected soils properties

Location	Clare	Tamworth	Hamilton
Soil Type	Neutral Clay loam	Alkaline Clay loam	Acid loam
pH (H ₂ O)	7.0	8.0	4.6
Organic C %	4.7	1.5	6.2
Nitrate-N mg/kg	7.6	65.3	93.0
Ammonium-N mg/kg	4.6	6.2	13.0

Results

Nitrification and N₂O

- ❖ Nitrification rates followed the order neutral clay loam < acid loam < alkaline clay loam (Table. 2).
- ❖ DMPP and C₂H₂ were most effective in the neutral clay loam (Table 2).
- ❖ C₂H₂ is more effective than DMPP in all three soils.
- ❖ DMPP and C₂H₂ can reduce N₂O emission flux from all three soils (Fig. 2).
- ❖ DMPP was less effective than C₂H₂ on N₂O emissions in the acid loam (Fig. 2, Table. 3).

Table 2. Nitrification rate (µg/g d⁻¹) and inhibition (%)^a by DMPP and C₂H₂ at day 28

Soil	Nitrification rate			Nitrification inhibition	
	Control	DMPP	C ₂ H ₂	DMPP	C ₂ H ₂
Neutral clay loam	1.7	1.1	0.5	81.2	84.1
Alkaline clay loam	5.7	3.8	0	52.8	62.4
Acid loam	3.2	1.9	0	42.3	62.6

^a % inhibition of nitrification = ((NO₃-N produced in control treatment)-(NO₃-N produced in inhibitor-treated soil))/(NO₃-N produced in control soil) × 100

Table 3. Mineral N concentration and cumulative N₂O emission at day 14^b

Soils	[NH ₄ ⁺ -N] (µg/g soil)			[NO ₃ ⁻ -N] (µg/g soil)			N ₂ O-N cumulative emission (g/ha)			N ₂ O/Nitrification ^c (%)	
	Control	DMPP	C ₂ H ₂	Control	DMPP	C ₂ H ₂	Control	DMPP	C ₂ H ₂	DMPP	C ₂ H ₂
Neutral Clay loam	49.0	73.0	82.2	37.8	11.1	9.2	3.4	0.6	0.8	25.2	28.3
Alkaline Clay loam	13.8	74.7	97.4	158.9	80.8	66.8	4.4	1.5	1.2	3.6	4.8
Acid loam	81.6	82.2	119.0	71.5	55.2	34.9	12.7	5.1	1.2	13.8	33.0

^b 14 days was the final collection time after 72 hrs closure (day 12 sample)

^c % N₂O/Nitrification = ((N₂O-N produced by nitrification over incubation period)/(NO₃-N produced over incubation period) × 100

Gene Abundance

- ❖ All three soils had detectable functional *amoA* gene from bacteria and archaea (Fig. 3).

Future work

- ❖ Quantification of the specific functional gene (*amoA*) to test for relationship between these nitrifiers and soil properties.

Gas collection vial

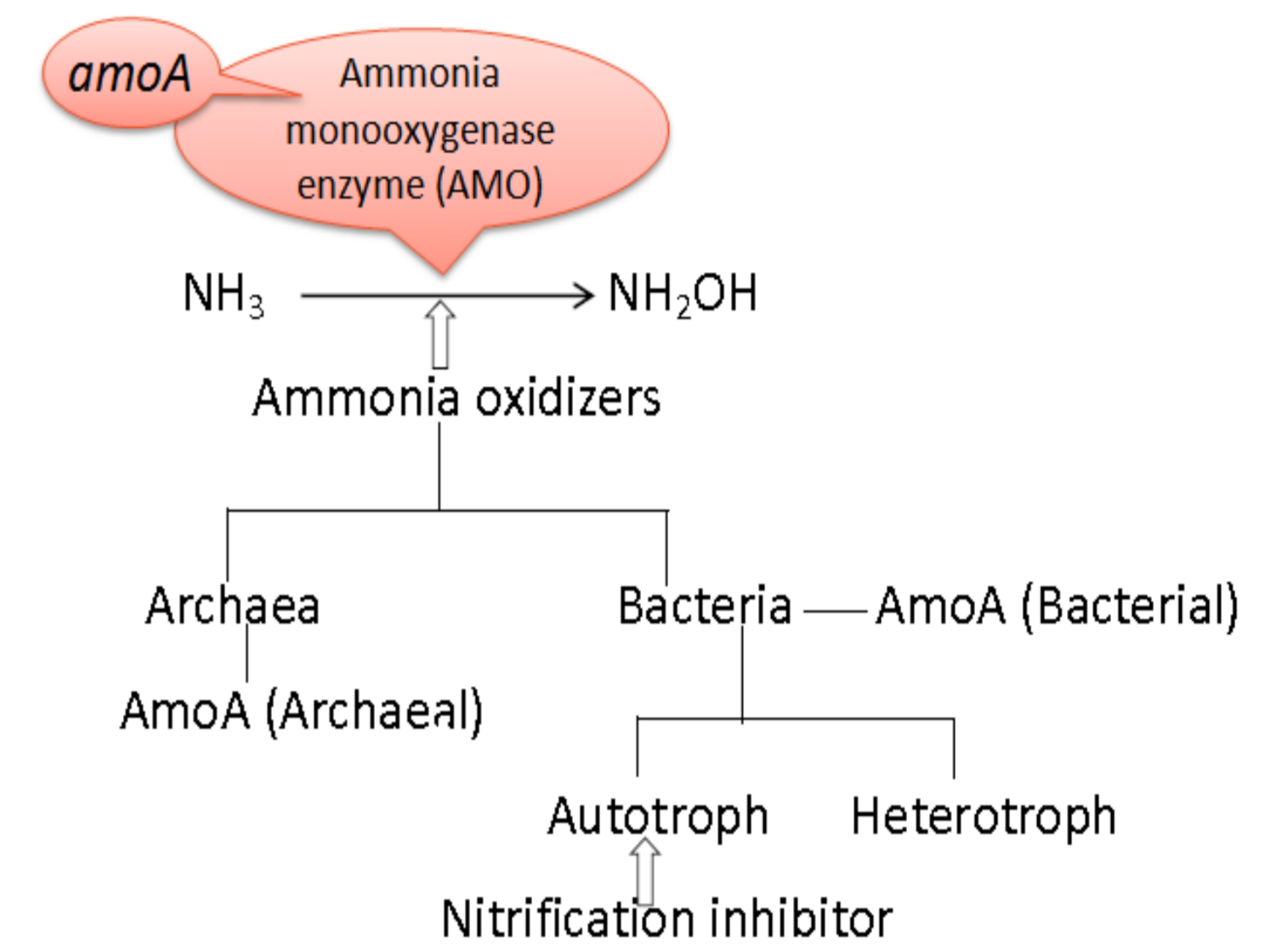
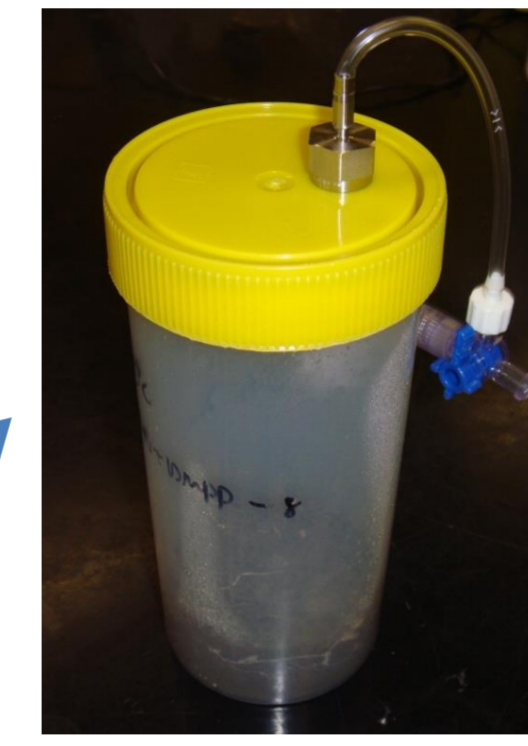


Fig. 1 Using functional *amoA* gene to separate archaea and bacteria

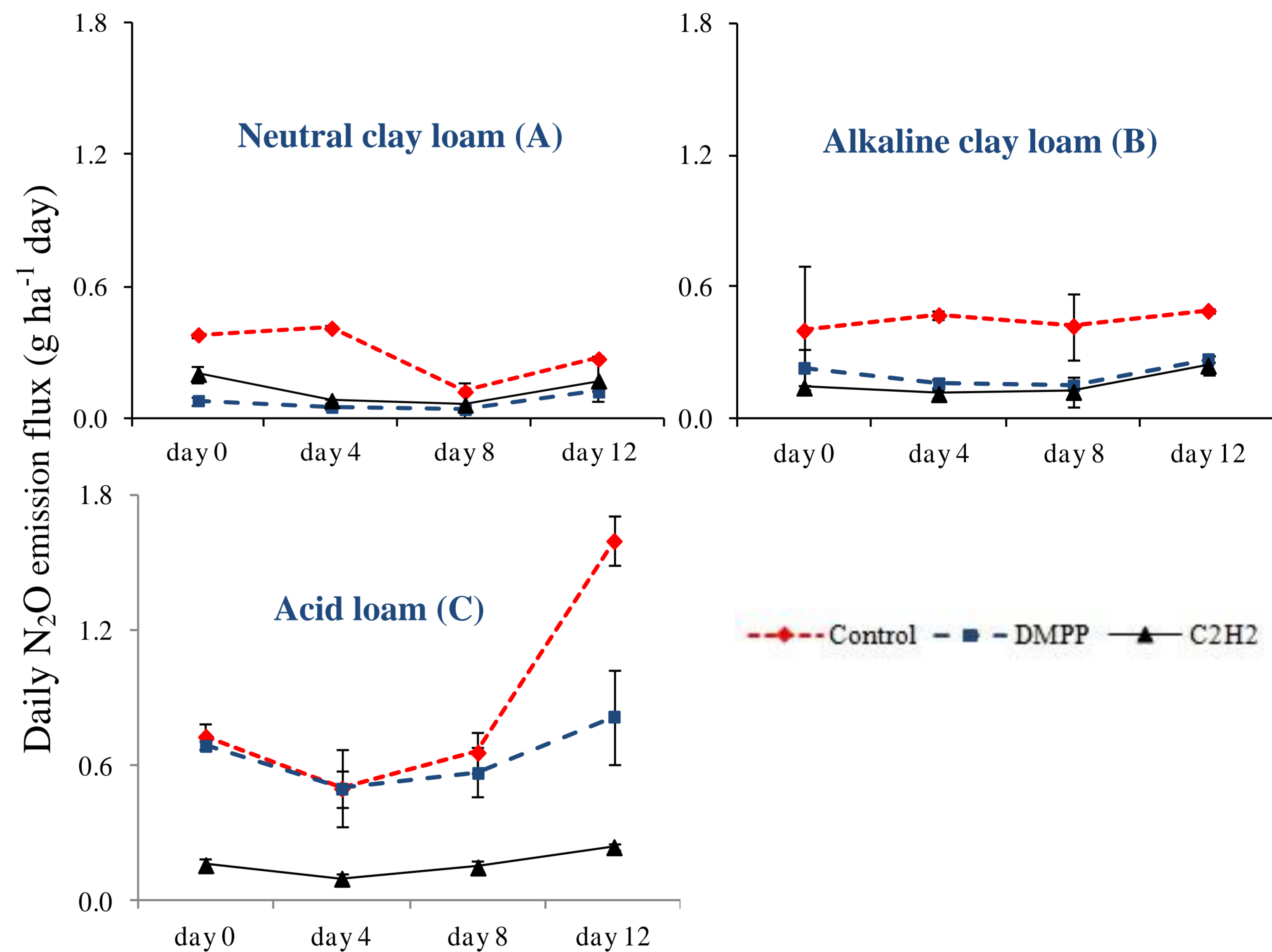


Fig. 2 Daily N₂O flux at 25°C. Error bars indicate standard deviations of four replicates.

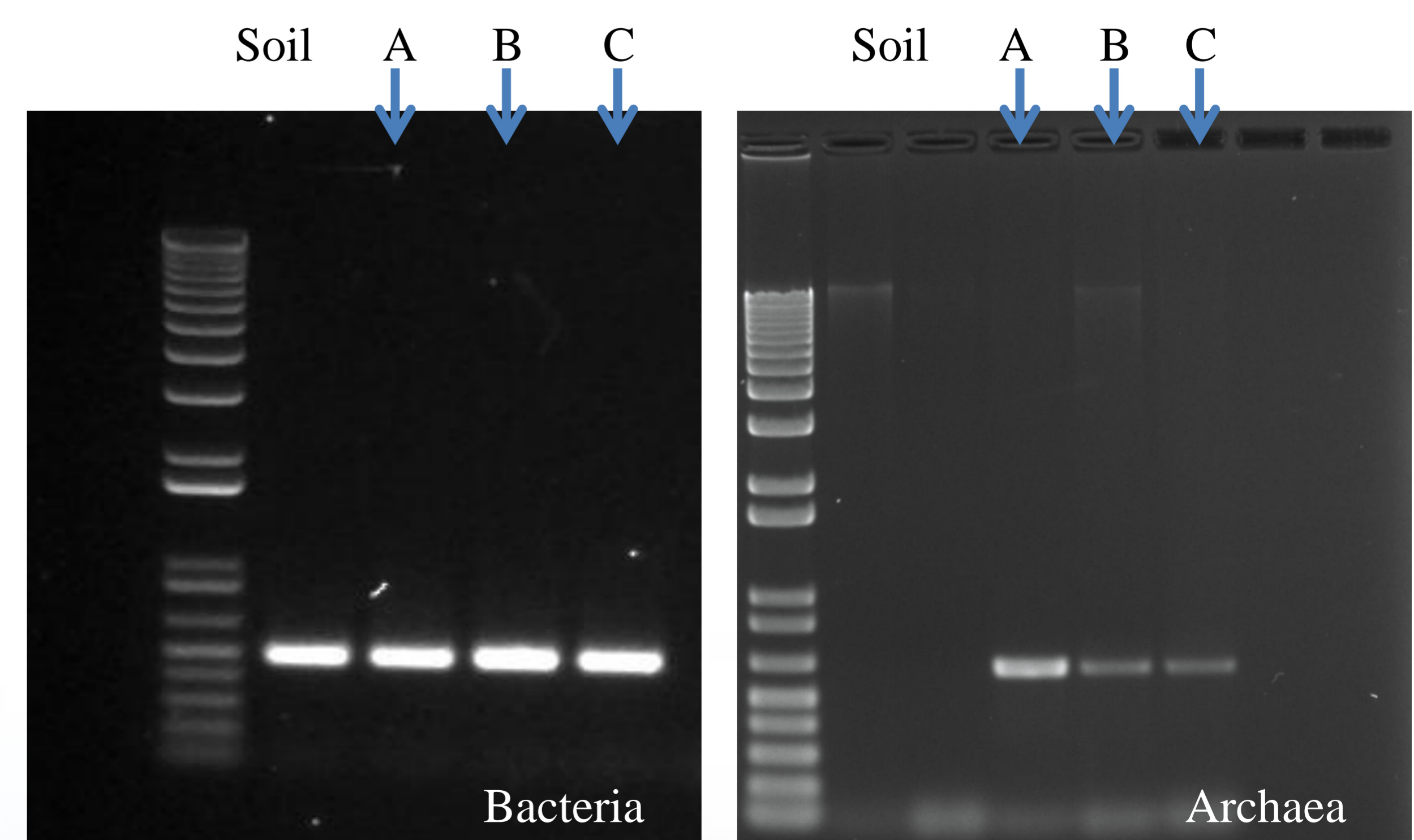


Fig. 3 Gel images showing detection of *amoA* gene from bacteria and archaea from PCR product